

COMPOSITION OF THE LEAF OILS OF *Clausena heptaphylla* AND *Clausena suffruticosa**

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Leaf oils obtained from *Clausena heptaphylla* and *Clausena suffruticosa*, were analyzed by GC-MS. Thirty six terpene components were identified in the *Clausena heptaphylla* oil. The main components were 3-carene (64.33%), α -phellandrene (5.74%) and l-4-terpineol (5.15%). Twenty two terpene components were identified in *Clausena suffruticosa* oil with the main components being esdragole (58.23%), anethole (33.20%), linalol (3.38%) and β -ocimene (1.40%).

Clausena heptaphylla (Roxb.) Wight & Arn. (Bengali name-“Karanphal”) is a shrub and *C. suffruticosa* Wight & Arn. (Bengali name-“Kalomorichal”) is an under shrub. Both the plants distributed in south and Southeast Asia and belong to the family Rutaceae (Guillaumin, 1911; Hooker, 1875). The plants of this genus are known to be useful in paralysis, ulcerated nose, colic, stomach trouble, fever and headache, muscular pain, malarial fever (Thuy *et al.*, 1999; Yusuf *et al.*, 1994). They are also reputed as diuretic, astringent, insecticide, tonic and vermifuge (Perry, 1980). The leaves of the plants possess antimicrobial properties (Sohrab *et al.*, 2001; Begum *et al.*, 2006). Previous phytochemical studies on *Clausena* led to the isolation of carbazole alkaloids (Thuy *et al.*, 1999), clausmarin (Sohrab, 2000), lunamarin C (Sohrab 2002), coumarins (Huang *et al.*, 1997) and limonoids (Ngadjui *et al.*, 1989). A new carbazole alkaloid, named clausenal, was isolated from the leaves of *C. heptaphylla* and identified as 1, 8-dimethoxy-3-formylcarbazole from physical, chemical and synthetic evidences. The alkaloid was found to be active against both Gram-positive and Gram-negative bacteria, and fung (Chakraborty *et al.*, 1995). In this paper, we report the results obtained by GC-MS on *C. heptaphylla* and *C. suffruticosa* leaf oils. This is the first report of the components of the leaf oils of *C. heptaphylla* and *C. suffruticosa*.

Fresh leaves of *C. heptaphylla* and *C. suffruticosa* were collected from the plants grown in the campus of BCSIR Laboratory, Chittagong during June 2007. Two-voucher specimens (Y-1006 and Y-1007) were deposited in the herbarium of BCSIR Laboratory, Chittagong, Bangladesh. The sundried leaves of *C. heptaphylla* and *C. suffruticosa* were cut into small pieces, and subjected to hydrodistillation method using Clevenger's apparatus for 4 hrs. The oil was extracted with diethyl ether and dried over anhydrous sodium sulfate.

GC-MS analysis: The essential oils from leaf of *C. heptaphylla* and *C. suffruticosa* were analyzed by GC-MS electron impact ionization (EI) method on GC-17A gas chromatograph (Shimadzu) coupled to a GC-MS QP 5050A Mass Spectrometer (Shimadzu); fused silica capillary column (30m x 2.5mm; 0.25 μ m film thickness), coated with DB-1 (J&W); column temperature 100°C (2 min) to 250°C at the rate of 3°C/min; carrier gas, helium at constant pressure of 90Kpa. Acquisition parameters full scan; scan range 40-350 amu. Comparing with the NIST library data identified the compounds.

The leaves of *Clausena heptaphylla* and *C. suffruticosa* native to Bangladesh yielded 0.2% and 0.25% (Volume/fresh wt.) of essential oils respectively. Table 1 showed the relative percentage of components for the *C. heptaphylla* oil. 36 components were identified that represented about 99.27% of the total oil. The main components were 3-carene (64.33%), followed by α -phellandrene (5.74%) and l-4-terpineol (5.15%). Other notable compounds are γ -elemene (1.08%), β -pinene (3.39%), α -pinene (3.45%), α -terpinene (1.07%), caryophyllene (1.18%), germacrene D (1.75%), isothujol (1.29%), m-cymene (2.29%) and terpinene (2.12%).

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Table 1. Constituents of the leaf essential oil from *C. heptaphylla*.

Name of compounds	Percent content
α -Bourbonene	0.04
δ -Cadinene	0.06
α -Caryophyllene	0.20
δ -Elemene	0.07
γ -Elemene	1.08
α -Farnesene	0.58
α -Guaiane	0.11
α -Phellandrene	5.74
β -Pinene	3.39
α -Pinene	3.45
α -Terpinene	1.07
α -Thujene	0.54
2-Carene	0.73
3-Carene	64.33
4-Vinylguaiaicol	0.23
Camphene	0.15
Caryophyllene	1.18
Caryophyllene oxide	0.37
Cis-Piperitol	0.11
Crypton	0.19
Dimethoxydurene	0.07
Estragole	0.16
Germacrene D	1.75
Isotujol	1.29
L-4-Terpineol	5.15
Lanceol,cis	0.10
m-Cymene	2.29
Menthol	0.13
Methyl-2, 5-Octadecadiyonate	0.13
Nerolidyl acetate	0.13
Pseudolimonene	0.24
Spathulenol	0.66
Tau-Cadinol	0.15
Terpinene	2.12
Tumerone	0.64
Z-Ocimene	0.77

Table 2 showed the presence of 22 components that represented 99.46% of the total oil of *C. suffruticosa* leaf. The main components were estragole (58.23%), followed by anethole (33.20%), linalol (3.38%) and β -ocimene (1.40%). These results showed that the oils were complex mixture of numerous compounds; many of which were terpenes in nature and presented in trace amounts. It has been observed that there is great variation in the chemical composition of *C. heptaphylla* and *C. suffruticosa* 3-carene (64.33%) was the main component in *C. heptaphylla* but totally absent in *C. suffruticosa*.

Table 2. Constituents of the leaf essential oil from *C. suffruticosa*.

Name of the components	Percent content
α -Pinene	0.19
Sabinene	0.03
β -Myrcene	0.03
4-Hexen-1-ol, acetate	0.02
Limonene	0.02
β -Ocimene	1.40
1-Methylhexyl acetate	0.03
Linalol	3.38
2-Dodecen-4-yne, (E)-	0.02
Esdragole	58.23
Linalyl acetate	0.06
Anethole	33.20
γ -Amylbutyrolactone	0.38
Eugenol	0.69
Caryophyllene	0.87
α -Caryophyllene	0.10
Isohomogenol	0.08
γ -Elemene	0.30
./-.-trans-Nerolidol	0.13
δ -Undecalactone	0.17
tau.-Cadinol	0.07
Cubenol	0.05

On the basis of above results it may be suggested that *C. heptaphylla* and *C. subtaphylla*, growing widely in Bangladesh, can be utilized as a source for the isolation of natural 3-carene and esdragole respectively. This is the first report of this kind of analysis on the leaf oils of *C. heptaphylla* and *C. subtaphylla*.

REFERENCES

- Begum, R., M. S. Rahman, M. R. Haque, A. M. S. Chowdhury, M. A. .Rashid. 2006. Antimicrobial Activity and Cytotoxicity of *Clausena suffruticosa*. Dhaka Univ. J. Pharm. Sci. 5, 81-83.
- Chakraborty, A., C. Saha, G. Podder. B. K. Chowdhury and P. Bhattacharyya. 1995. Carbazole alkaloid with antimicrobial activity from *Clausena heptaphylla*. Phytochemistry. 38, 787-789.
- Guillaumin, A. 1911. Rutaceae. In: Lecomte, H. Flore générale de l'Indo-Chine 1, 629-687.
- Hooker, J. D. 1875. *Flora of British India*, Vol. I. Reeve & Co. Ltd, England. P. 506.
- Huang S. C., P. L. Wu and T. S. Wu. 1997. Two coumarins from the root bark of the *Clausena excavata*. Phytochemistry. 44, 179-181.
- Ngadjui, B. T., J. F. Ayafor and B. L..Sondengam. 1989. Limonoids from *Clausena anisata*. J. Nat. Prods. 52, 832-836.
- Perry, L. M. 1980. *Medicinal plants of East and Southeast Asia: attributed properties and uses*. Cambridge: The MIT Press. p. 363.
- Sohrab, M. H., C. M. Hasan and M. A. Rashid. 2002. Lunamarin C, a new terpenoid coumarin from *Clausena heptaphylla*. Pharmazie. 57, 573-574.
- Sohrab, M. H., C. M. Hasan and M. A. Rashid. 2000. Clausmarin-A from the leaves of *Clausena heptaphylla*. Biochemical Systematics and Ecology. 28, 91-93.
- Sohrab, M. H., M. A. Mazid, E. Rahman, C. M. Hasan and M. A. Rashid. 2001. Antibacterial activity of *Clausena heptaphylla*. Fitoterapia. 72, 547-549.
- Thuy, T. T., H. Ripperger, A. Porzel, T. V. Sung and G. Adam. 1999. Counlarins, limonoids and an alkaloid from *Clausena excavata*. Phytochemistry. 52, 511-516.
- Yusuf, M., J. U. Chowdhury, M. A. Wahab and J. Begum. 1994. Medicinal Plants of Bangladesh. 1st edition, Bangladesh Council of Scientific and Industrial Research (BCSIR), Dhaka.